The calibration curves were constructed with $1/x^2$ weighting and the regression coefficients were greater than 0.99.

2.7. Other analyses

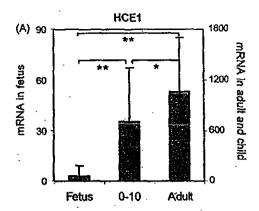
Protein concentrations were determined with BCA assay (Pierce) based on bovine serum albumin standard. Data are presented as mean ± S.D. All enzymatic assays were repeated three times with the same microsomal preparation. Statistical tests were performed to compare means (Student's t-test) or correlations (Spearman). In all cases, significant differences were assumed when p-values were less than 0.05.

3. Results

3.1. Ontogenic expression of human carboxylesterases HCE1 and HCE2 by RT-qPCR

In this study, we first evaluated the expression patterns of human carboxylesterases HCE1 and HCE2 in various age groups. A total of 104 RNA samples were analyzed including 48 samples from fetuses (82-224 gestation days), 34 from children (0 days-10 years of age) and 22 from adults (>18). The levels of HCE1 and HCE2 mRNA were determined with RT-qPCR, and the results are summarized in Fig. 1 and Table 2. Overall, the adult group had the highest levels of HCE1 and HCE2 mRNA, and the fetal group had the lowest levels for both enzymes. Based on the values of the means, the adult group expressed HCE1 at levels 319-fold higher than the fetal group, and ~50%. higher than the child group (Fig. 1 and Table 2). Likewise, the adult group expressed HCE2 at levels 55-fold higher than the fetal group and ~40% higher than the child group. In all cases, the differences among various age groups were statistically significant (Fig. 1). It should be noted that two different sets of-Y axial values were used in Fig. 1 to accommodate the large difference in the mRNA levels between the fetal and the other two groups.

In addition to the large difference among various age groups (inter-group), a large inter-individual variability was detected within a group. The fetal group, for example, showed a 431-fold difference (ratio between the maximum and the minimum) in HCE1 mRNA with a coefficient of variation (CV) of as high as 172% (Table 2). The child and adult groups, on the other hand, varied less in HCE1 mRNA with a 218- and 12-fold difference, respectively (Table 2). Similarly, the levels of HCE2 mRNA varied in all age groups, however, the overall variability was much less than HCE1 mRNA. The variation in HCE2 mRNA



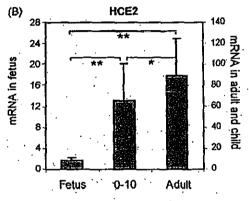


Fig. 1 – Levels of HCE1 and HCE 2 mRNA in the adult, child and fetal groups. Total RNAs were subjected to RT-qPCR analysis for the level of HCE1 mRNA (A) and HCE2 mRNA (B) by Taqman probes as described in Section 2. The adult group contained 22 samples, the child group contained 34 samples and the fetal group contained 48 samples. The signals from each target were normalized based on the signal from PolII and expressed as relative levels among all samples. The data are presented as mean \pm S.D. (*) Statistical significance at p < 0.001.

was the same in the fetal and child groups and higher than that in the adult group (21- versus 4-fold) (Table 2).

We next examined whether the expression of carboxylesterases is age-related within a group. The levels of mRNA were plotted against age or gestation days, and the correlation coefficients were computed. The adult group showed no clear correlation for either HCE1 or HCE2 (data not shown). In the fetal samples, the level of HCE2 but not HCE1 mRNA was

Group	n	Minimum	Maximum	Variability	Mean	S.D.	. CV (%)
Fetus-HCE1	48	0.07	30.15	431 (fold)	3.32	5.70	172
.Child-HCE1	34	12.18	2657.77	218	711.47	638.51	. 90
Adult-HCE1	.22	225.50	2659.61	12	.1059.59	644,92	61
Fetus-HCE2	48 .	0.20	4.28	21	1.63	1.36	. 9 i .
Child-HCE2	34	7.00	148.78	• 21 •	65.82	34.90	53
Adult-HCE2	22.	39.89	168.66	4.	89:87	34:83	₹ 3 9

Abbreviations: S.D.: standard deviation; CV: coefficient of variation.

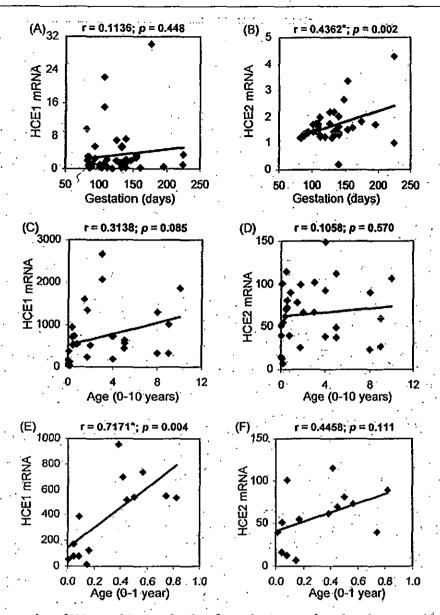


Fig. 2 – Age-related expression of HCE1 and HCE2. The data from Fig. 1 were plotted against age with SPSS version 16. (A) Correlation of HCE1 mRNA with gestation days. (B) Correlation of HCE2 mRNA with gestation days. (C) Correlation of HCE1 mRNA with age in the child group (0–10 years old). (D) Correlation of HCE2 mRNA with age in the child group (0–10 years old). (E) Correlation of HCE1 mRNA with age in the sub-child group (0–1-year-old). (F) Correlation of HCE2 mRNA with age in the sub-child group (0–1-year-old), (f) statistical significance (p < 0.05).

significantly correlated with age (Fig. 2A and B). In contrast, the child group showed better correlation with age on HCE1 than HCE2 mRNA, although neither correlation reached the level of statistical significance (Fig. 2C and D). To gain additional information on age-related expression, correlation analysis was performed on the samples from donors under one year old (0–1 year). In this sub-group, much improved correlation was observed on both HCE1 and HCE2 with a p-value of 0.004 and 0.111, respectively (Fig. 2E and F).

3.2. Inter-individual variability in carboxylesterase proteins and oseltamivir hydrolysis in the child group

We next examined whether the levels of carboxylesterase mRNA reflected the levels of corresponding proteins. RNA-

microsome matched samples from 11 available pediatric-aged subjects were evaluated. Fetal matched samples were not tested because of low mRNA expression and low activities towards marker substrates (described below). No RNA-microsome matched samples were available for the adult group. The microsomal samples were first analyzed by Western immunoblotting with antibody against HCE1 or HCE2, and the immunostaining intensities were quantified by the KODAK 1D Image Analysis Software. As shown in Fig. 3A, all samples contained HCE1 and HCE2 proteins, but the relative abundance varied markedly. In particular, one donor (lane 6) showed extremely low levels of both carboxylesterases (Fig. 3A). Based on the immunostaining intensities, HCE1 protein varied by ~100-fold, and HCE2 protein varied by ~20-fold. The inter-individual variability, however, was decreased

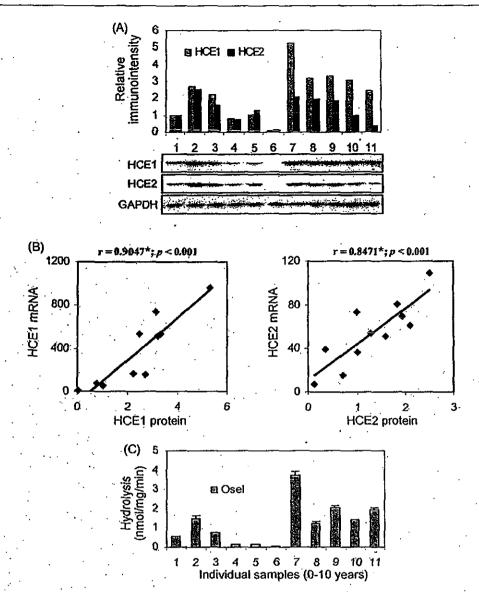


Fig. 3 – Individual variation of HCE1 and HCE2 proteins and oseltamivir hydrolysis in the child group. (A) Western analyses. Microsomes (1.5 μ g) were resolved by 7.5% SDS-PAGE and transferred electrophoretically to nitrocellulose membranes. The blots were incubated with an antibody against HCE1, HCE2 or GAPDH and chemiluminescent substrate. The signal was captured by a KODAK Image Station 2000 and the relative intensities were quantified by the KODAK 1D Image Analysis Software. (B) Correlation analyses. The immunointensities of HCE1 and HCE2 were plotted against the levels of respective mRNA. The correlation coefficients and evaluations on statistical significance were performed with SPSS version 16. (1) Statistical significant (p < 0.001). (C) Oseltamivir hydrolysis by individual liver samples of the child group. Microsomes (20 μ g) were incubated with oseltamivir (200 μ M) at 37 °C for 10 min, and the formation of oseltamivir carboxylate was detected by LC-MS/MS. Data were assembled from three independent experiments with two injections of each experiment.

to \sim 8-fold for both carboxylesterases when the data-point (lane 6) was eliminated. For both HCE1 and HCE2, the mRNA levels were correlated significantly with the levels of respective proteins (p < 0.001) (Fig. 3B). The HCE1 matched samples had a correlation coefficient of 0.9047, and the HCE2 matched samples had a slightly lower correlation coefficient (0.8471) (Fig. 3B).

The large individual variability in carboxylesterase protein pointed to the possibility of marked differences in the metabolism of therapeutic agents and other xenobiotics. To directly test this possibility, the anti-influenza viral agent

oseltamivir (an ester prodrug) was incubated with individual donor samples (Fig. 3A), and the hydrolysis was monitored. As shown in Fig. 3C, all samples hydrolyzed this anti-viral agent, and the overall hydrolysis varied by 127-fold. Such a large inter-individual variability was in agreement with the variation in the abundance of HCE1 (Fig. 3A), which has been shown to catalyze the hydrolysis of oseltamivir (3). As expected, sample 6 (lane 6) contained the lowest HCE1 protein and showed the lowest hydrolytic activity toward oseltamivir. Conversely, sample 7 (lane 7) contained the highest level of HCE1 protein and was the most active toward this anti-viral

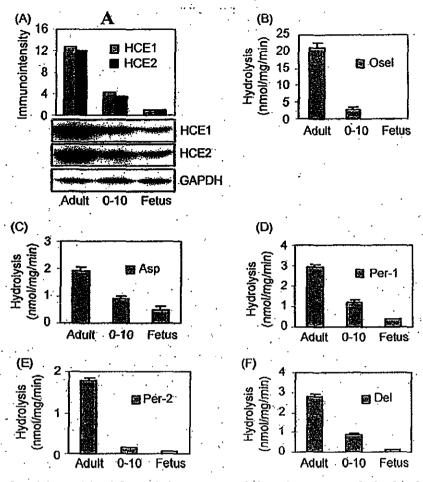


Fig. 4 – Hydrolysis of oseltamivir, aspirin, deltamethrin and permethrin. (A) Western analysis of pooled samples for various groups. Microsomes (1.5 µg) pooled from the adult, child and fetal group were analyzed by Western blotting and the immunostaining intensities were quantified by the KODAK 1D Image Analysis Software. (B) Hydrolysis of oseltamivir by pooled microsomes. Microsomes (20 µg) from various age groups were incubated with oseltamivir (200 µM) at 37 °C for 10 min, and the formation of oseltamivir carboxylate was detected by LC-MS/MS. (C) Hydrolysis of aspirin by pooled microsomes. Microsomes (80 µg) from various age groups were incubated with aspirin (1 mM) at 37 °C for 60 min, and the formation of salicylic acid was detected by HPLC. (D and E) Hydrolysis of permethrin by pooled microsomes. Microsomes (50 µg) from various age groups were incubated with permethrin (100 µM) at 37 °C for 60 min, and the disappearance of the parent compounds was detected by HPLC. Permethrin contained a mixture of cis- and trans-isomers at a ratio of 46 and 52%, respectively with the cis-form having a retention time of 10.41 and the trans-form of 10.18. (F) Hydrolysis of deltamethrin by pooled microsomes. Microsomes (50 µg) from various age groups were incubated with deltamethrin (100 µM) at 37 °C for 60 min, and the disappearance of the parent compound was detected by HPLC. Data were assembled from three independent experiments with each experiment having two injections.

agent (Fig. 3A and C). The hydrolysis of oseltamivir among these individual samples was highly correlated with the protein level of HCE1 with a correlation coefficient of 0.9373, although samples 8 and 10 exhibited relatively lower hydrolysis compared with their relatively HCE1 contents (Fig. 3A and C).

3.3. Hydrolysis of drugs and insecticides by liver microsomes of fetuses, children and adults

We next extended the metabolism study to include the antiinflammatory agent aspirin, and insecticides permethrin (cisand trans-) and deltamethrin. In addition to their pharmacological and toxicological implication, these chemicals were chosen because they are hydrolyzed in an isoform-specific manner. Oseltamivir and deltamethrin are predominately hydrolyzed by HCE1 [3,19,20], whereas aspirin is predominately hydrolyzed by HCE2 [4]. The cis-form of permethrin is favorably hydrolyzed by HCE2, whereas the trans-form is comparably hydrolyzed by both forms. The extended metabolism study was performed with microsomes pooled from various age groups, and the microsomes were so pooled by mixing equal amount from all individuals in an age group. The pooled samples enabled comparison to be made on the overall hydrolysis of these chemicals among different age groups.

We first determined the levels of HCE1 and HCE2 in the pooled samples by Western immunoblotting. Based on the immunostaining intensities, the child group expressed ~25%

of carboxylesterases (both HCE1 and HCE2) of the adult group and the fetal group expressed less than 10% of the adult group (Fig. 4A). Overall, the magnitude of hydrolysis of these chemicals was correlated well with the relative levels of carboxylesterases in these samples (Fig. 4B-F). The adult pooled sample showed the highest activity toward all chemicals and the fetal pooled samples showed the lowest activity. Hydrolysis of aspirin and trans-permethrin (Per-1) by the samples pooled from the fetuses and children was slightly higher than that predicted according to their relative abundance of carboxylesterases. For example, the fetal sample contained less than 10% of carboxylesterases of the adult sample but showed 25% of aspirin hydrolysis (Fig. 4A and C). Conversely, the hydrolysis of oseltamivir, cis-permethrin and deltamethrin by the fetal group was less than 10% of the adult group. The precise mechanism on these discrepancies remains to be determined. On the other hand, the higher-than predicted hydrolysis, in the case of aspirin, was likely due to hydrolysis by other enzymes (highly expressed in the fetal liver) or due to polymorphic variants. In support of these possibilities, butyrylcholinesterase has been shown to hydrolyze aspirin [21], and certain polymorphic variants of HCE2 were found to differ from the wild-type enzyme in hydrolyzing this anti-platelet agent [4]. In addition, the liver expresses a third carboxylesterase [22], but it remains to be determined whether this carboxylesterase hydrolyzes these compounds (i.e., aspirin and trans-permethrin) and whether the expression of this carboxylesterase is developmentally regulated.

4. Discussion

Carboxylesterases constitute a class of hydrolytic enzymes that play important roles in the metabolism of therapeutic. agents and detoxication of insecticides [1]. In this study, we analyzed a large number of individual liver samples for the expression patterns of HCE1 and HCE2, two human carboxylesterases predominately expressed in the liver. Overall, the adult group expressed significantly higher HCE1 and HCE2 than the child group or the fetal group. The age-related expression was confirmed on the levels of both mRNA and protein. In agreement with the expression patterns, the adult microsomes were approximately 4 times as active as the child microsomes and more than 10 times as active as the fetal microsomes in hydrolyzing a group of therapeutic agents and insecticides. Even within the same age group, a large interindividual variability was detected in mRNA and protein levels as well as hydrolytic activity.

Although both HCE1 and HCE2 exhibited a similar expression pattern among the various age groups, there were several major differences. First, HCE1 exhibited much greater intergroup and inter-individual variability than HCE2. For example, based on the values of the means, the adult group displayed a 319-fold higher level in HCE1 mRNA compared with the fetal group. In contrast, these two groups showed only a 55-fold difference in the level of HCE2 mRNA (Fig. 1 and Table 2). Likewise, the adult group showed a 430-fold inter-individual variability in HCE1 mRNA (ratio between maximum over minimum), in contrast, only a 21-fold difference in HCE2 mRNA was detected in the same group (Table 2). Second, HCE1

_ 73.53...

displayed better age-related expression than HCE2 in the child group with correlation coefficients of 0.3138 and 0.1058, respectively (Fig. 2C and D). In contrast, HCE2 displayed better correlation in the fetal group with correlation coefficients of 0.4362 and 0.1136, respectively (Fig. 2A and B).

The large inter-individual variability, particularly in the fetal and children groups, was likely an outcome coordinated by multiple mechanisms. In this study, we have shown that the adult group expressed the highest levels of HCE1 and HCE2 followed by the child group, and the fetal group expressed the lowest levels of both enzymes (Fig. 1 and Table 2). Such agerelated expression patterns were confirmed by RT-qPCR and Western analyses (Figs. 1, 3 and 4) and established that developmental regulation is involved in the expression of HCE1 and HCE2. However, the correlation with age in many cases was only moderate at the most and did not reach the levels of statistical significance (Fig. 2C and F). Although the precise mechanisms remain to be determined, the lack of strong correlation with age in these groups was likely due to complicated factors such as the administration of therapeutic agents and disease conditions. We have previously reported that pathological condition and therapeutic agents markedly altered the expression of HCE1 and HCE2 [6,8], Interleukin-6, a cytokine usually elevated during inflammation, profoundly suppressed the expression of both HCE1 and HCE2 (8). Consistent with the suppression of carboxylesterases by cytokines, patients with elevated cytokine conditions such as liver cirrhosis had much lower capacity of hydrolyzing ester drugs such as perindopril, a non-sulphydryl angiotensin converting enzyme inhibitor [23,24].

The significantly lower level of HCE1 in the child group. compared with the adult group, provides a molecular explanation to the large pharmacokinetic difference in oseltamivir between these two groups. In this study, the sample pooled from the children was only ~15% as active as the sample pooled from the adults in hydrolyzing oseltamivir (Fig. 4C). Consistent with the in vitro metabolism, children under 12 years old reportedly produced only approximately half of the hydrolytic metabolite produced by adults [25]. Apparently the low level production of oseltamivir carboxylate. in children was likely due to ineffective hydrolysis of the parent drug and higher clearance of the metabolite. Furthermore, we have shown that individual samples in the child group varied by as many as 127-fold in oseltamivir hydrolysis (Fig. 4A). Pharmacokinetic studies in children, however, did not detect such a large inter-individual variation in the production of hydrolytic metabolite of oseltamivir [25,26]. One explanation is that the frequency with an extremely low expression level of HCE1 is rare in the general population, and the pharmacokinetic studies were performed in 24 or fewer children [25,26]. Indeed, there was a reported rare case that an adult patient with diabetes mellitus had only 1-2% capacity of normal people in hydrolyzing clopidogrel based on the values of the means and standard deviations [27]. Like oseltamivir, clopidogrel is a substrate of HGE1 [3]. Ineffective hydrolysis of oseltamivir, on the other hand, likely leads to increased concentration in the brain. Some patients taking oseltamivir reportedly developed neurobehavioral changes [28], although a direct link remains to be established between the developed neurotoxiciy and the use of oseltamivir.

In contrast to oseltamivir, pyrethroids have long been recognized to exert neurotoxicity [29]. As a class of the most used insecticides in the world, both the general population and workers have a high risk to be exposed to these insecticides. Epidemiological studies have shown that the exposure level, in some cases, can be high [30,31]. Pyrethroid insecticides are generally considered safe to mammals, because they are rapidly eliminated by carboxylesterases. In this study, we have shown that the fetuses and children hydrolyzed pyrethroids at a rate of only ~20% or lower of the adults (Fig. 4D-F), suggesting their vulnerability to pyrethroids-induced toxicity. In support of this notion, neonatal rats were reportedly 17 times as sensitive as adult rats to cypermethrin [12]. Neurotoxicity induced by pyrethroids appears to cause irreversible damage. Prenatal exposure to deltamethrin, for example, led to a deficit in locomotor activity of offspring post-natally at 9 weeks [32]. In humans, micromolar concentrations were reported in the meconium [33]. This is particularly of relevance as fetuses have only limited capacity of hydrolytic detoxication as described in this report.

In summary, our work points to several important conclusions. First, the expression of both HCE1 and HCE2 increases with age, establishing that their expression is developmentally regulated and that fetuses and children generally have lower capacity of hydrolytic metabolism than adults. Second, there is a large inter-individual variability in the expression of these enzymes, particularly in the fetal and child groups. It is likely that the expression of HCE1 and HCE2 in these age groups is subjected to non-developmental regulation with high sensitivity (e.g., xenobiotic regulation). Carboxylesterases are recognized to play important roles in drug metabolism and insecticide detoxication. The findings on the large variability among different age groups or even within the same age group have important pharmacological and toxicological implications, particularly in relation to altered pharmacokinetics of ester drugs in children and vulnerability of fetuses and children to insecticides such as pyrethroids.

REFERENCES

- Satoh T, Hosokawa M. Structure, function and regulation of carboxylesterases. Chem Biol Interact 2006;162:195-211.
- [2] Schwer H, Langmann T, Daig R, Becker A, Aslanidis C, Schmitz G. Molecular cloning and characterization of a novel putative carboxylesterase, present in human intestine and liver. Biochem Biophys Res Commun 1997;233:117–20.
- [3] Shi D, Yang J, Yang D, LeCluyse EL, Black C, You L, et al. Anti-influenza prodrug oseltamivir is activated by carboxylesterase human carboxylesterase 1, and the activation is inhibited by antiplatelet agent clopidogrel. J Pharmacol Exp Ther 2006;319:1477-84.
- [4] Tang M, Mukundan M, Yang J, Charpentier N, LeCluyse EL, Black C, et al. Antiplatelet agents aspirin and clopidogrel are hydrolyzed by distinct carboxylesterases, and clopidogrel is transesterificated in the presence of ethyl alcohol. J Pharmacol Exp Ther 2006;319:1467–76.
- [5] Wu MH, Yan B, Humerickhouse R, Dolan ME. Irinotecan activation by human carboxylesterases in colorectal adenocarcinoma cells. Clin Cancer Res 2002;8:2696–700.

- [6] Zhu W, Song L, Zhang H, Matoney L, LeCluyse E, Yan B. Dexamethasone differentially regulates expression of carboxylesterase genes in humans and rats. Drug Metab Dispos 2000;28:186-91.
- [7] Morgan EW, Yan B, Greenway D, Parkinson A. Regulation of two rat liver microsomal carboxylesterase isozymes: species differences, tissue distribution and the effects of age, sex and xenobiotic treatment of rats. Arch Biochem Biophys 1994;315:514-26.
- [8] Yang J, Shi D, Yang D, Song X, Yan B. Interleukin-6 suppresses the expression of carboxylesterases HCE1 and HCE2 through transcriptional repression. Mol Pharmacol 2007;72:686-94.
- [9] Oxford JS, Mann A, Lambkin R. A designer drug against influenza: the NA inhibitor oseltamivir (Tamiflu). Expert Rev Anti Infect Ther 2003;1:337-42.
- [10] Anand SS, Kim KB, Padilla S, Muralidhara S, Kim HJ, Fisher JW, et al. Ontogeny of hepatic and plasma metabolism of deltamethrin in vitro: role in age-dependent acute neurotoxicity. Drug Metab Dispos 2006;34:389–97.
- [11] Pope CN, Chakraborti TK, Chapman ML, Farrar JD, Arthun D. Comparison of in vivo cholinesterase inhibition in neonatal and adult rats by three organophosphorothicate insecticides. Toxicology 1991;68:51-61.
- [12] Cantalamessa F. Acute toxicity of two pyrethroids, permethrin, and cypermethrin in neonatal and adult rats. Arch Toxicol 1993;67:510-3.
- [13] Sheets LP, Doherty JD, Law MW, Reiter LW, Crofton KM. Age-dependent differences in the susceptibility of rats to deltamethrin. Toxicol Appl Pharmacol 1994;126:186-90.
- [14] Pope CN, Karanth S, Liu J, Yan B. Comparative carboxylesterase activities in infant and adult liver and their in vitro sensitivity to chlorpyrifos oxon. Regul Toxicol Pharm 2005;42:62-9.
- [15] Vyhlidal CA, Gaedigk R, Leeder JS. Nuclear receptor expression in fetal and pediatric liver: correlation with CYP3A expression. Drug Metab Dispos 2006;34:131-7.
- [16] Wortham M, Czerwinski M, He L, Parkinson A, Wan YJ. Expression of constitutive androstane receptor, hepatic nuclear factor 4 alpha, and P450 oxidoreductase genes determines interindividual variability in basal expression and activity of a broad scope of xenobiotic metabolism genes in the human liver. Drug Metab Dispos 2007;35: 1700-10.
- [17] Leeder JS, Gaedigk R, Marcucci KA, Gaedigk A, Vyhlidal CA, Schindel BP, et al. Variability of CYP3A7 expression in human fetal liver. J Pharmacol Exp Ther 2005;314:626–35.
- [18] Radonić A, Thulke S, Mackay IM, Landt O, Siegert W, Nitsche A. Guideline to reference gene selection for quantitative real-time PCR. Biochem Biophys Res Commun 2004;313:856-62.
- [19] Godin SJ, Scollon EJ, Hughes MF, Potter PM, DeVito MJ, Ross MK. Species differences in the in vitro metabolism of deltamethrin and esfenvalerate: differential oxidative and hydrolytic metabolism by humans and rats. Drug Metab Dispos 2006;34:1764-71.
- [20] Nishi K, Huang H, Kamita SG, Kim IH, Morisseau C. Hammock BD Characterization of pyrethroid hydrolysis by the human liver carboxylesterases hCE-1 and hCE-2. Arch Biochem Biophys 2006;445:115-23.
- [21] Kolarich D, Weber A, Pabst M, Stadlmann J, Teschner W, Ehrlich H, et al. Glycoproteomic characterization of butyrylcholinesterase from human plasma. Proteomics 2008;8:254–63.
- [22] Sanghani SP, Quinney SK, Fredenburg TB, Davis WI, Murry DJ, Bosron WF. Hydrolysis of irinotecan and its oxidative metabolites, 7-ethyl-10-[4-N-(5-aminopentanoic acid)-1piperidino] carbonyloxycamptothecin and 7-ethyl-10-[4-(1piperidino)-1-amino]-carbonyloxycamptothecin, by human

- carboxylesterases CES1A1, CES2, and a newly expressed carboxylesterase isoenzyme, CES3. Drug Metab Dispos 2004;32:505–11.
- [23] Thiollet M, Funck-Brentano C, Grange JD, Midavaine M, Resplandy G, Jaillon P. The pharmacokinetics of perindopril in patients with liver cirrhosis. Br J Clin Pharmacol 1992;33:326-8.
- [24] Eriksson AS, Gretzer C, Wallerstedt S. Elevation of cytokines in peritoneal fluid and blood in patients with liver cirrhosis. Hepatogastroenterology 2004;51:505-9.
- [25] Oo C, Barrett J, Hill G, Mann J, Dorr A, Dutkowski R, et al. Pharmacokinetics and dosage recommendations for an oseltamivir oral suspension for the treatment of influenza in children. Paediatr Drugs 2001;3:229–36.
- [26] Oo C, Hill G, Dorr A, Liu B, Boellner S, Ward P. Pharmacokinetics of anti-influenza prodrug oseltamivir in children aged 1-5 years. Eur J Clin Pharmacol 2003;59:411-5.
- [27] Heestermans AA, van Werkum JW, Schömig E, ten Berg JM, Taubert D. Clopidogrel resistance caused by a failure to metabolize clopidogrel into its metabolites. J Thromb Haemost 2006;4:1143-5.

- [28] FDA Patient Safety News (2007) Caution on Neuropsychiatric Events with Tamiflu: Show #59:http:// www.accessdata.fda.gov/psn/printer.cfm?id=486.
- [29] Ray DE, Fry JR. A reassessment of the neurotoxicity of pyrethroid insecticides. Pharmacol Ther 2006;111:174-93.
- [30] Leng G, Kühn KH, Idel H. Biological monitoring of pyrethroids in blood and pyrethroid metabolites in urine: applications and limitations. Sci Total Environ 1997;199:173–81.
- [31] Heudorf U, Angerer J. Metabolites of pyrethroid insecticides in urine specimens: current exposure in an urban population in Germany. Environ Health Perspect 2001;109:213-7.
- [32] Johri A, Yadav S, Singh RL, Dhawan A, Ali M, Parmar D. Long lasting effects of prenatal exposure to deltamethrin on cerebral and hepatic cytochrome P450 s and behavioral activity in rat offspring. Eur J Pharmacol 2006;544:58-68.
- [33] Ostrea Jr EM, Bielawski DM, Posecion Jr NG, Corrion M, Villanueva-Uy E, Jin Y, et al. A comparison of infant hair, cord blood and meconium analysis to detect fetal exposure to environmental pesticides. Environ Res 2008;106:277–83.



異常行動モデルとしての薬物誘発ジャンピング行動に対するタミフルの 影響とその防御法に関する研究

〇小野信文、牛島逸子、木村公彦 福岡大・薬・医薬品情報学

【背景と目的】 タミフルを代表とする抗インフルエンザウイルス剤服用者が、異常行動の結果、事故死を起こしたことが報告され、緊急安全性情報が出された。その後この因果関係に関しては国家的研究班が組織され、親在研究が行われているが、詳細は未だ明らかでない。しかし、世界的に見れば新たなインフルエンザウイルスの出現もあり、それらの対処は急がねばならない状況と思われる。このような状況を踏まえ、我々は新たな視点から抗インフルエンザウイルス薬の欠点を締う可能性を検討した。

【実験方法】ジャンピング行動は、直径32cmの正人角形、高さ35cmの台に置いたマウスが薬物接与後40分間に下の床に飛び降りる行動とし、飛び降りる主での時間と回数を(飛び降りた動物は台に戻し)概定した。尚、マウスは測定までこの測定台へ置かれたことはない。薬物の投与スケジュールは、haloperidol 0.5 mg/kg (ip) 5 分後に clonidine 10mg/kg (ip) 投与し、測定台へ移し制定を開始した。タミフルの1%CMC 経濁被は haloperidol 16 分前に経口投与し、前処置薬はタミフルのさらに10 分前にio 投与した。

【結果並びに考察】タミフル150mg/kg、haloperidol、clonidine それぞれ単独投与では制定時間内にジャンピング行動は全く起こさなかった。haloperidol と clonidine の併用では、0.63 回座のジャンピング行動が見られた。この haloperidol clonidine 防発行動は、タミフル150mg/kg の併用により22.6 回/匹と有意に増強された。このタミフルによる増強作用は、acetazolamide 20 mg/kg 前処匿により 0.50 回/匹と有意に抑制し、150 mg/kg 前処匿ではジャンピング行動を完全に消失させた。さらに、diazepam 0.5、1 mg/kg、valprorate 40 mg/kg、fluoxetine 5 mg/kg 前処匿により、タミフルによる薬物誘発ジャンピング行動は消失した。したがって、このようなジャンピング行動は、異常行動の一つの指標として有用と考えられる。現時点ではこれらの行動の発現機序は不明であるが、各種中枢神経伝達物質系の不均衡が関与し、抑制作用はその是正によることが示唆される。

A-06

Morphine 誘発精神依存形成における L 型高電位開口性カルシウムチャネル (HVCCs) 機能亢進に対する PI 3-kinase の関与

〇芝崎真裕、黒川和宏、桂 昌司、大能誠太郎 川崎医大・薬理学

【目的】我々は既に、精神依存の評価法である条件づけ場所培好性試験において、L型 HVCC 拮抗薬である nifedipine により、morphine 誘発報酬効果が有意に抑制されることを報告した。一方、PL 8-kinase class III である Vps34は trafficking に深く関与することが報告さていることがら、morphine による L型 HVCC の発現増加に関与している可能性が考えられる。そこで本研究では、inorphine による精神依存形成過程における L型 HVCC の発現増加を伴った機能亢進機序を、精神依存マウスおよび依存性薬物を連続爆露した初代培養大脳皮質神経細胞(神経細胞)を用いて、行動薬理学的および神経科学的観点から検討した。

【方法】Morphineによる報酬効果は条件づけ場所管好性試験により行った。神経細胞へのmorphine の連続環路は、Hanks 被で希釈したものを直接培養液中に添加した。【**Gaz*引流入は2分間の30 mM KCI 刺激により神経細胞内へ取り込まれた放射活性を測定した。蛋白差現最は Western blot 法により、解析した。

【結果および考察】Morphine による報酬効果は、L型HVCC阻害薬(nifedipine)の前処置により完全に消失した。この時点での倒坐核を含む領域および大脳皮質画分におけるL型HVCCaleおよびa218 subunit ならびに Vps34 蛋白の発現量に有意な増加が認められた。同様に、神経細胞に morphine を連続機器した場合に視察される 80 mM KCl 誘発[45Ca**]流入の増加は、L型HVCC 阻害薬の併用により完全に阻害された。また、morphine の連続曝露により L型HVCCaleおよびa218 subunit ならびに Vps34 蛋白量の有意な増加が認められた。このale subunit 蛋白量の増加は、PI 3 kinase 阻害薬(LY294002)の併用処置により有意に減少した。以上の成績より、morphine によるL型HVCCale subunit 蛋白発現増加機構に、Vps34 が一部関与することが明らかとなった。

日本菜理学会近畿部会,第113回(2008.06.20,岡山)