Table 3

Extent of TSE agent removal during nanofilitation of plasma-derived coagulation factor concentrates. Adapted from Refs. [43,62]

	•	_		-		
Starting plasma fraction	Nanofilter	End-product	Agent	Spike	Assays	Reduction factor
DEAE-Toyopearl 650M eluate	Planova 35N	vWF	263K	MF	WB ·	≥ 3.1 [61]
DEAE-Toyopearl 650M eluate	Pianova 35N + 15N	FVIII	263K	MF	Bioassay	≥ 3.3 [61]
SD + DEAE-Toyopearl 650M eluate	Planova 35N + 15N	FVIII	263K	MF	WB .	≥ 5.1 [61]
Monoclonal antibody chromatography eluate	Planova 75N + 35N + 35 + 15N	FVIII	263K.	BH + SD	Bioassay	4 + 0.3 + 1.3 +≥ 0.5 [62]
Monoclonal antibody chromatography eluate	Planova 75N + 35N + 35N + 15N	FVIII	263K	PrPsc + SD	Bioassay	$3.1 + 0 + 0.8 + \ge 2$ [62]
Ion-exchange chromatography + heparin-Sepharose eluate	Planova 15N	FIX	263K	MF	Bioassay	4.8 [61]
Ion-exchange chromatography + heparin-Sepharose eluate	Planova 15N	FIX	BSE 6PB1	BH + sonication	Bioassay	5.3 [42]

MF: microsomal fraction; BH: brain homogeneate; WB: Western-blot; SD: solvent-detergent.

tate [39]. Using blood from scrapie-infected hamsters, and in spiking studies where scrapie 263K PrPsc was added to human plasma, 20% and 10% of the infectivity partitioned in the cryoprecipitate, respectively [35,54]. By contrast, when human plasma was spiked with scrapie 263K brain homogeneate (BH), 90% of PrPsc was found in the cryoprecipitate. These contradictory results may highlight the influence of the nature of the experimental model used, in particular the physicochemical nature of the spike [34], or of the variations in the down-scaling of the cryoprecipitation procedure, or they actually illustrate the fact that cryoprecipitation does not ensure robust partitioning of TSE infectivity.

3.3.2.2. Ethanol precipitation. The capacity of precipitation steps to remove prions efficiently has first been shown by partitioning of endogenous infectivity using a rodent model. It is particularly well documented for the ethanol fractionation process isolating albumin and immunoglobulins. In the albumin fractionation procedure using either the Cohn-Oncley or the Kistler and Nitchman processes, major and consistent reduc-

tion factors (typically 3-5 logs) of TSE agent have been found by various groups, most specifically during the precipitation of fraction II + III, fraction III, and fraction IV, or their equivalents using slightly modified fractionation conditions [35,42] (Table 4). Similar experiments revealed 3-5 logs removal during the precipitation III or I+III used in the IgG process [35,42]. These data suggest that, in spite of variations in the conditions (such as ethanol concentration and pH) used, reproducible clearance of PrP^{TSE} takes place. Removal is achieved when the precipitate is separated. It has been speculated that prion removal in precipitates is possibly due to aggregation and is dependent upon pH and presence of alcohol [35,55]. Other precipitation steps, using caprylic acid during immunoglobulins production [56] or polyethylene glycol also contribute to prion removal [42].

3.3.2.3. Depth filtration. Depth filters are made of a combination of a matrix (generally based on cellulose), filter aids (diatomeous earth, resins, or other adsorbents), and a drainage system. They are used to clarify crude protein solutions and

Table 4
Extent of TSE agent removal during the ethanol plasma fractionation process in the manufacture of albumin and immunoglobulin G. Adapted from Refs. [41,43,62]

Step	Fraction evaluated	Agent	Spike	Assays	Reduction factor (log ₁₀)
Precipitation of fraction I	Supernatant	263K	вн	WB	1.1 [41,43]
Precipitation of I + II + III	-	263K	MF	WB	1.3 [34]
Precipitation of I + II + III	-	263K	· MF	WB	≥ 2.8° [42]
Precipitation of fraction II + III	_	263K	вн	WB	≥ 4.7 [41,43]
_	_ '	263K	вн	Bioassay	- 6.0 [41]
_ `	_	Sc237	BH/MF/CLD/PrPSc	CDI	3.6/3.1/3.1/4.0 [34]
_	_	263K	BH	WB	≥4.2/≥4.1 [41,43]
	~	263K	вн	Bioassay	3.7/4.6 [41]
Precipitation of fraction I + III'	-	263K	MF	WB	≥ 3.5° [42]
Precipitation of fraction IV	_	263K	MF	WB	≥ 3.0 [35]
_	-	Sc237	BH/MF/CLD/PrPSc	CDI	3.2/3.4/3.2/2.2 [34]
_	-	263K	MF	WB	≥ 3.7 [35]
_	_	263K	MF	WB	≥ 4.3" [42]
-	-	263K	BH .	WB	≥ 4.3 [41,43]
_	_	263K	BH	Bioassay	5.3 [41]

BH: brain homogeneate; MF: microsomal fraction; CLD: caveolae-like domain; WB: Western-blot; CDI: conformation-dependent immunoassay.

^{*} Evaluated together with filter aids to remove precipitates.

b Precipitate discarded during the manufacture of IgG.

^e Precipitate discarded during the manufacture of albumin.

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Table 5

Extent of TSE agent removal during depth filtration of albumin and immunoglobulin G fractions. Adapted from Refs. [41,43,62]

Fraction	Filter	Agent	Spike	Assays	Reduction factor (log ₁₀)
Supernatant I	Seitz Supra P80	Sc237	BH/MF	CDI	-0.1/0.1 [34]
Supernatant III	Millipore AP20	BSE 301V	MF	Bioassay	2.4 [37]
	Seitz KS80P	BSE 301V	MF	Bioassay	≥3.1 [37]
_	Cuno Zetaplus	263K	вн	WB	≥3.3 [55]
Supernatant IV	Seitz Supra P80	Sc237 -	CLD/PrPsc	CDI	≥ 0.9/ ≥ 2.4 [34]
	Seitz AKS5 (carbon)	263K	MF	WB	2.7 [42]
Fraction V (albumin)	Cuno Delipid-I	263K	MF	WB	23 (35)
	Seitz KS80P	263K	MF	WB	≥4.9 [35]
Fraction II (IgG)	Seitz K200P	263K	MF	WB	≥ 2.8 [35]
-	Ca ₃ PO ₄ + filter aid + Cuno	263K	MF	Bioassay	2.5 [42]
_	Cuno	263K	вн	Bioassay	≥4.9 (57)

BH: brain homogeneate; MF: microsomal fraction; CLD: caveolae-like domains; CDI: conformation-dependent immunoassay; WB: Western-blot.

Table 6
Extent of TSE agent removal during nanofiltration of plasma-derived albumin, and IgG. Adapted from Refs. [43,62]

End-product	Nanofilter	Agent	Spike	Assays	Reduction factor
Albumin ^a	Planova 35N	CID	· BH	Bioassay	≥5.9 [53]
Albumin* .	Planova 35N	ME7	вн	Bioassay	4.93 [36]
Albumin + detergent	Planova 35N	ME7	вн	Bioassay	1.61 [36]
Albumin*	Planova 15N	ME7	вн	Bioassay	≥ 5.87 [36]
Albumin + detergent	Planova 15N	ME7	вн	Bioassay	≥4.21 [36]
RhO (D) IgG	VIRESOLVE 180	263K	Detergent treated, sonicated	WB	≥ 2.5 [63]
	•		and filtered BH		·
IgG	DV50	263K	BH	Bioassay	4.4 [64]
IgG	Planova 75N + 35N	263K	MF	Bioassay	3.2 [61]

BH: brain homogeneate: MF: microsomal fraction: WB: Western-blot.

remove precipitates. As such they are an important adjunct to the ethanol precipitation process. Principle of action-encompasses both removal of particulates larger than the pore-size by size-exclusion, and of smaller elements by adsorption. During immunoglobulin manufacture, the supernatant of Fraction III (Supernatant III) and the re-dissolved Fraction II precipitate, and during production of albumin, the Supernatant IV and the re-dissolved Fraction V, generally undergo depth filtration steps. Experimental spiking experiments (Table 5) have shown that several types and grades of depth filters can remove prions [35,37,42]. Depending upon the type of depth filter or the physico-chemical parameters of the suspension, PrPTSE removal may be due to an aggregation in the presence of alcohol [55] or to hydrophobic adsorption on the filter aids [42]. The impact of protein composition and content remains to be investigated and understood to demonstrate and guarantee the robustness of this non-specific removal.

3.3.2.4. Nanofiltration. Table 6 shows experimental data on the removal of TSE agents during nanofiltration of albumin and IgG. As for coagulation factors, high reduction factors have been found. Prion partitioning is presumed to result from a size-exclusion mechanism.

3.4. Cleaning and sanitization

Most equipment, including stainless steel reactors, chromatographic gels and columns, ultrafilters are re-used and, therefore, must undergo steam (SIP) or chemical (CIP) sanitizing procedures between production batches. These processes

should be validated to ensure proper bacterial, pyrogenic, viral, and protein decontaminations. Experience gained with the sterilization of surgical instruments has shown that autoclaving at 134 °C for 18 min or 121 °C for 30 min reduced the transmission of prion infectivity by a factor > 5 log₁₀ [57, 58] but autoclaving without immersion is less effective (4-4.5 log reduction) [58]. Standard chemical decontamination methods (NaOH 1 N, NaOCl 20,000 ppm) and hydrogen peroxide alone achieved a reduction of > 6.5 and 4.5 log₁₀, respectively [58]. By experiments involving a hamster scrapie strain 263K BH model, it was shown that 0.1 M NaOH for 15 min, in the absence of detergent, at 4 and 18 °C caused a reduction of 3.5 and 4.0 log₁₀ of the prion protein, respectively. In the presence of sarkosyl, a 60-min incubation in NaOH further enhanced PrPRES reduction to > or $= 4.5 \log_{10}$, with no residual infectivity. Therefore 0.1 N NaOH could also effectively inactivate prions, and its efficacy can be enhanced by the addition of sarkosyl [59]. A separate study shows that 0.1 M NaOH at 60 °C for 2 min and 0.25 M NaOH at 30 °C for 60 min inactivate 3.96 and 3.93 log₁₀ of mouse-adapted scrapie strain ME7, respectively, and 0.5 M NaOH at 30 °C for 60 or 75 min inactivates \geq 4.23 and 4.15 \log_{10} [60].

4. Conclusion

There is so far no evidence of transmission of prions by plasma derivatives. The prevalence of the disease in the population is considered to be very low, although possibly not quite as low as initially considered [5,6]. Based on experimental models, it is believed that the infectivity in the plasma does

^{*} Nanofiltration is not used during production of albumin preparations; a detergent was added for experimental purposes only.

not exceed a few infectious doses per ml. By lack of knowledge of the nature of the agent associated to the infectivity in plasma, and in the absence of validated screening tests, alternative precautionary measures have been introduced to prevent the possibility of transmission of vCJD by plasma derivatives. Epidemiological surveillance of the population is in place in countries where BSE and vCJD cases have been identified. In some countries, blood donation deferral criteria include travel or residence of donors in BSE countries, and history of previous transfusion or tissue transplantation. Based on filtration experiments of blood collected from scrapie-infected hamsters, leucoreduction decreases by about 50% the prion infectivity in plasma [22,27]. Extent of removal of TSE agents during the plasma fractionation process appears to be substantial. Data from various laboratories and using different experimental models show several logs removal of TSE infectivity during the fractionation process. The most effective, but nonspecific, removal steps are ethanol precipitation, depth filtration, and ion-exchange chromatography. Nanofiltration was also demonstrated to remove several logs of TSE infectivity, possibly based on a specific prion removal mechanism by size-exclusion. Uncertainty on the validity of these experimental studies remains, and additional studies are needed, since the biochemical features of the infective agent in blood and plasma is not known, nor the extent to which it may be present in blood donations. Research should continue, aiming at identifying the features of TSE agents in human plasma and at ensuring the robustness of prion removal steps and sanitizing procedures during plasma product manufacture.

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References

- Burnouf T. Modern plasma fractionation. Transfus Med Rev 2007 (in press).
- [2] Llewelyn CA, Hewitt PE, Knight RS, Amar K, Cousens S, Mackenzie J, et al. Possible transmission of variant Creutzfeldt-Jakob disease by blood transfusion. Lancet 2004;363:417-21.
- [3] Hewitt PE, Llewelyn CA, Mackenzie J, Will RG. Creutzfeldt-Jakob discase and blood transfusion: results of the UK Transfusion Medicine Epidemiological Review study. Vox Sang 2006;91:221-30.
- [4] WHO. WHO guidelines on tissue infectivity distribution in transmissible spongiform encephalopathies. http://www.who.int/bloodproducts. 2006.
- [5] Hilton DA, Ghani AC, Conyers L, Edwards P, McCardle L, Ritchie D, et al. Prevalence of lymphoreticular prion protein accumulation in UK tissue samples. J Pathol 2004;203:733-9.
- [6] Ironside JW, Bishop MT, Connolly K, Hegazy D, Lowrie S, Le Grice M, et al. Variant Creutzfeldt-Jakob disease: prion protein genotype analysis of positive appendix tissue samples from a retrospective prevalence study. BMJ 2006;332:1186-8.
- [7] WHO. Guidelines on viral inactivation and removal procedures intended to assure the viral safety of human blood plasma products. www.WHO. int. Geneva; 2003. 1-72 p.

- [8] WHO. Recommendations for the production, quality control and regulation of plasma for fractionation. http://www.who.int/bloodproducts. 2005.
- [9] Aguzzi A, Heikenwalder M. Pathogenesis of prion diseases: current status and future outlook. Nat Rev Microbiol 2006;4:765-75.
- [10] Cervia JS, Sowemimo-Coker SO, Ortolano GA, Wilkins K, Schaffer J, Wortham ST. An overview of prion biology and the role of blood filtration in reducing the risk of transfusion-transmitted variant Creutzfeldt-Jakob disease. Transfus Med Rev 2006:20:190-206.
- [11] Foster PR. Assessment of the potential of plasma fractionation processes to remove causative agents of transmissible spongiform encephalopathy. Transfus Med 1999;9:3-14.
- [12] Silveira JR, Raymond GJ, Hughson AG, Race RE, Sim VL, Hayes SF, et al. The most infectious prion protein particles. Nature 2005;437:257-61.
- [13] Burnouf T, Radosevich M. Reducing the risk of infection from plasma products: specific preventative strategies, Blood Rev 2000;14:94-110.
- [14] Taylor DM. Inactivation of transmissible degenerative encephalopathy agents: a review. Vet J 2000;159:10-7.
- 15] Prowse C. Prion removal with filters, ISBT Science Series 2006;1:230-4.
- [16] Cervenakova L, Yakovleva O, McKenzie C, Kolchinsky S, McShane L, Drohan WN, et al. Similar levels of infectivity in the blood of mice infected with human-derived vCJD and GSS strains of transmissible spongiform encephalopathy. Transfusion 2003;43:1687-94.
- [17] Gregori L, Lambert BC, Gurgel PV, Gheorghiu L, Edwardson P, Lathrop JT, et al. Reduction of transmissible spongiform encephalopathy infectivity from human red blood cells with prion protein affinity ligands. Transfusion 2006;46:1152-61.
- [18] Houston F, Foster JD, Chong A, Hunter N, Bostock CJ. Transmission of BSE by blood transfusion in sheep. Lancet 2000;356:999-1000.
- [19] Mathiason CK, Powers JG, Dahmes SJ, Osborn DA, Miller KV, Warren RJ, et al. Infectious prions in the saliva and blood of deer with chronic wasting disease. Science 2006;314:133-6.
- [20] Saa P, Castilla J, Soto C. Presymptomatic detection of prions in blood. Science 2006;313:92-4.
- [21] Beringue V, Benesik A, Le Dur A, Reine F, Lai TL, Chenais N, et al. Isolation from cattle of a prion strain distinct from that causing bovine spongiform encephalopathy. PLoS Pathog 2006;10:2.
- [22] Prowse C. Controlling the blood-borne spread of human prion disease. ISBT Science Series 2006;1:21-4.
- [23] Afssaps. Analyse du risque de transmission de la variante de la maladie de Creutzfeldt-Jakob par les médicaments d'origine humaine et par les produits sanguins labiles---actualisation des données du rapport du groupe ad hoc de décembre 2000. Saint-Denis, France: Afssaps; rapport de mars 2003 (1-22 p).
- [24] Chabanel A, Sensebe I, Masse M, Maurel JP, Plante J, Hivet D, et al. Quality assessment of seven types of fresh-frozen plasma leucoreduced by specific plasma filtration. Vox Sang 2003;84:308-17.
- [25] Bumouf T, Kappelsberger C, Frank K, Burkhardt T. Residual cell content in plasma from 3 centrifugal apheresis procedures. Transfusion 2003; 31:1522-6.
- [26] Klein MA, Frigg R, Flechsig E, Raeber AJ, Kalinke U, Bluethmann H, et al. A crucial role for B cells in neuroinvasive scrapic. Nature 1997; 390:687-90.
- [27] Gregori L, McCombie N, Palmer D, Birch P, Sowemimo-Coker SO, Giulivi A, et al. Effectiveness of leucoreduction for removal of infectivity of transmissible spongiform encephalopathies from blood. Lancet 2004;364: 529-31
- [28] Brown P, Cervenakova L, McShane LM, Barber P, Rubenstein R, Drohan WN. Further studies of blood infectivity in an experimental model of transmissible spongiform encephalopathy, with an explanation of why blood components do not transmit Creutzfeldt-Jakob disease in humans. Transfusion 1999;39:1169-78.
- [29] Prowse CV, Bailey A. Validation of prion removal by leucocytedepleting filters: a cautionary tale. Vox Sang 2000;79:248.
- [30] Miura M, Nirasawa H, Yamashita E, Kobayashi K, Inadome S, Dieter J, et al. Evaluation of a new combination filter for prion and leukoreduction (LR) of red cell concentrates (RCC). Transfusion 2006;46(supplement): 109A (Abstract SP221).

- [31] Burnouf T. Chromatography in plasma fractionation: benefits and future trends. J Chromatogr B Biomed Appl 1995;664:3-15.
- [32] Burnouf T, Radosevich M. Affinity chromatography in the industrial purification of plasma proteins for therapeutic use. J Biochem Biophys Methods 2001;49:575-86.
- [33] CPMP. Note for guidance on virus validation studies: the design, contribution and interpretation of studies validating the inactivation and removal of viruses (revised). CPMP/BWP/CPMP/5136/03. http://www.emea.eu.int. London: European Agency for the Evaluation of Medicinal Products (EMEA); 1996.
- [34] Vey M, Baron H, Weimer T, Grouer A. Purity of spiking agent affects partitioning of prions in plasma protein purification. Biologicals 2002;30: 187-96.
- [35] Foster PR, Welch AG, McLean C, Griffin BD, Hardy JC, Bartley A, et al. Studies on the removal of abnormal prion protein by processes used in the manufacture of human plasma products. Vox Sang 2000;78: 86-95.
- [36] Tateishi J, Kitamoto T, Mohri S, Satoh S, Sato T, Shepherd A, et al. Scrapie removal using Planova virus removal filters. Biologicals 2001; 29:17-25.
- [37] Reichl HE, Foster PR, Welch AG, Li Q, MacGregor IR, Somerville RA, et al. Studies on the removal of a bovine spongiform encephalopathy-derived agent by processes used in the manufacture of human immunoglobulin. Vox Sang 2002;83:137-45.
- [38] Stenland CJ, Lee DC, Brown P, Petteway Jr. SR, Rubenstein R. Partitioning of human and sheep forms of the pathogenic prion protein during the purification of therapeutic proteins from human plasma. Transfusion 2002;42:1497-500.
- [39] Brown P, Rohwer RG, Dunstan BC, MacAuley C, Gajdusek DC, Drohan WN. The distribution of infectivity in blood components and plasma derivatives in experimental models of transmissible spongiform encephalopathy. Transfusion 1998;38:810-6.
- [40] Foster PR. Removal of TSE agents from blood products. Vox Sang 2004;87(Suppl 2):7-10.
- [41] Lee DC, Stenland CJ, Miller JL, Cai K, Ford EK, Gilligan KJ, et al. A direct relationship between the partitioning of the pathogenic prion protein and transmissible spongiform encephalopathy infectivity during the purification of plasma proteins. Transfusion 2001;41:449-55.
- [42] Flan B, Aubin JT. Évaluation de l'efficacité des procédés de purification des proteins plasmatiques à éliminer les agents transmissibles non conventionnels. Virologie 2005;9:S45-56.
- [43] Lee DC, Stenland CJ, Hartwell RC, Ford EK, Cai K, Miller JL, et al. Monitoring plasma processing steps with a sensitive Western blot assay for the detection of the prion protein. J Virol Methods 2000;84:77-89.
- [44] Flan B. Tissue culture system for TSE detection in process studies. 2006 April 11-12. 2006; Paris: International Plasma Fractionation Association.
- [45] Saa P, Castilla J, Soto C. Ultra-efficient replication of infectious prions by automated protein misfolding cyclic amplification. J Biol Chem 2006; 46:35245-52.
- [46] Burnouf T, Burnouf-Radosevich M, Huart JJ, Goudemand M. A highly purified factor VIII:c concentrate prepared from cryoprecipitate by ionexchange chromatography. Vox Sang 1991;60:8-15.
- [47] Burnouf T, Michalski C, Goudemand M, Huart JJ. Properties of a highly purified human plasma factor IX:e therapeutic concentrate prepared by conventional chromatography. Vox Sang 1989;57:225-32.
- [48] Foster PR, Griffin BD, Bienek C, McIntosh RV, MacGregor IR, Somerville RA, et al. Distribution of a bovine spongiform encephalopathy-

- derived agent over ion-exchange chromatography used in the preparation of concentrates of fibrinogen and factor VIII. Vox Sang 2004;86:92-9.
- [49] Flan B. Validation de l'efficacité des étapes de fractionnement du plasma dans l'élimination des ATNC. Sang Thrombose Vaisseaux 2001;13:20– 8.
- [50] Porte P, Aubin JT, Arrabal S, Kimmel-Jehan C, Chtourou S, Flan B. Solvent-detergent treatment and ion-exchange chromatography have no impact on prion removal by 15 nm nanofiltration in the manufacture of Factane. Blood (American Society of Hematology Annual Meeting Abstracts). Blood 2005;106:4177.
- [51] Burnouf T, Radosevich M, Goubran HA, Willkommen H. Place of nanofiltration for assuring viral safety of biologicals. Current Nanoscience 2005;1:189-201.
- [52] Burnouf T, Radosevich M. Nanofiltration of plasma-derived biopharmaceutical products. Haemophilia 2003;9:24-37.
- [53] Tateishi J, Kitamoto T, Ishikawa G, Manabe S. Removal of causative agent of Creutzfeldt-Jakob disease (CJD) through membrane filtration method. Membrane 1993;18:357-62.
- [54] Rohwer RG. Experimental studies of blood infected with TSE agents. Proceedings of the Fourth Meeting of the FDA Advisory Committee on Transmissible Spongiform Encephalopathics. (Bethesda, USA); 1998 (p. 46-65).
- [55] Van Holten RW, Autenrieth SM. Evaluation of depth filtration to remove prion challenge from an immune globulin preparation. Vox Sang 2003; 85:20-4
- [56] Trejo SR, Hotta JA, Lebing W, Stenland C, Storms RE, Lee DC, et al. Evaluation of virus and prion reduction in a new intravenous immunoglobulin manufacturing process. Vox Sang 2003;84:176–87.
- [57] Vadrot C, Darbord JC. Quantitative evaluation of prior inactivation comparing steam sterilization and chemical sterilants: proposed method for test standardization. J Hosp Infect 2006;64:143-8.
- [58] Fichet G, Comoy E, Duval C, Antloga K, Dehen C, Charbonnier A, et al. Novel methods for disinfection of prion-contaminated medical devices. Lancet 2004;364:521-6.
- [59] Bauman PA, Lawrence LA, Biesert L, Dichtelmuller H, Fabbrizzi F, Gajardo R, et al. Critical factors influencing prion inactivation by sodium hydroxide. Vox Sang 2006;91:34-40.
- [60] Unal A, Thyer J, Uren E, Middleton D, Braun M, Maher D. Investigation by bioassay of the efficacy of sodium hydroxide treatment on the inactivation of mouse-adapted scrapic. Biologicals 2006 (in press).
- [61] Flan B. Evaluation of TSE removal procedures in the manufacture of plasma products. WHO Consultation on Tissue Infectivity Distribution in TSEs. 2005 14-16 September. 2005; Geneva: World Health Organization.
- [62] Losikoff A. Retention of TSE infectivity by Planova nanofilters as function of spike composition. In: IBC USA Conference Prions: Assessment and management of risks from blood-borne TSE infectivity. 2001 April 2. 2001; Westborough, USA: IBC USA Conference.
- [63] Van Holten RW, Autenrieth S, Boose JA, Hsieh WT, Dolan S. Removal of prion challenge from an immune globulin preparation by use of a sizeexclusion filter. Transfusion 2002;42:999-1004.
- [64] Gregori L, Maring JA, MacAuley C, Dunston B, Rentsch M, Kempf C, et al. Partitioning of TSE infectivity during ethanol fractionation of human plasma. Biologicals 2004;32:1-10.

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Subject PRO/AH/EDR> Arenavirus, organ transplants - Australia (VIC)

ARENAVIRUS, ORGAN TRANSPLANTS - AUSTRALIA (VICTORIA)

A ProMED-mail post

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[1]

Date: Sun 22 Apr 2007

Source: Herald Sun online [edited]

<http://www.news.com.au/sundayheraldsun/story/0,,21598166-2862,00.html</p>

Australia: Novel virus responsible for deaths of organ donation recipi

A virus unknown to medical science was behind the deaths of 3 Victorians who received organs from the one donor. The unnamed bug has been linked to Ebola virus, [a virus] responsible for the deaths of thousands in central Africa since the 1970s. [This is an incorrect statement. The organ transplant-associated virus is not related to Ebola virus; see part [2] below. - Mod.CP]. After baffling local scientists, experts from New York's Columbia University were called in to help solve the mystery of the multiple transplant deaths being investigated by the coroner.

Initial investigations and tests had been unable to determine any common link between the donor and the 3 recipients. The presence of the virus in the recipients is thought to be a world first. One of the New York team said: "The discovery of this virus is of national and international significance."

The Sunday Herald Sun revealed the deaths in February 2007. A 63-year-old woman died after receiving a kidney transplant at Austin Hospital. A 64-year-old man died after receiving a liver transplant there. The 3rd victim received a kidney at Royal Melbourne Hospital.

The male donor whose organs carried the suspected killer bug had died in Dandenong Hospital of a brain hemorrhage in December 2006 after returning from overseas; it is believed most of his trip was spent in Europe.

The virus is part of the rodent-borne arenavirus family and can cause "old-world" diseases such as Yellow Fever, Ebola and Lymphocytic choriomeningitis. [This statement is incorrect: yellow fever is caused by a flavivirus and Ebola hemorrhagic fever is caused by a filovirus; only lymphocytic choriomeningitis (LCM) is caused by an old-world arenavirus. - Mod.CP]. Victoria's acting Chief Health Officer, Dr John Carnie, confirmed the virus [LCM virus?] had been detected in multiple samples from all 3 transplant patients. But there was no evidence the virus represented a public health risk, he said.

Health authorities are examining whether future donated organs can be screened for [LCM?] virus. A spokesman for the Victoria Coroner's office said families of the victims were told yesterday [21 Apr 2007]. There would be a formal inquest.

Experts from Columbia's Greene Infectious Diseases Laboratory helped

solve the mystery. Initial investigations and tests were unable to determine any common link between the donor and the 3 recipients. Dr Carnie said the risk to the public was minimal because "these viruses [?] affect immunocompromised people, and it is rarely fatal in those with normal immune systems. We have not had any indication of any unexplained illnesses among families of the donor or recipients," he said. "This would be the case if it was transmissible person to person. Our supposition is it was transmitted by organ transplantation."

Cutting edge techniques were used for the 1st time by the Greene lab -- in collaboration with Victorian Infectious Diseases Reference Laboratory -- to gene sequence the virus. "Our gene technology enables unbiased sequencing of all agents present," Columbia's Prof. Ian Lipkin said. "We found a handful (of combinations) that were related to Lassa virus or LCM virus [both old world arenaviruses -- Mod.CP]. Using these clues we can confidently say this is a new virus, present in the original organs and so different than anything seen before."

Communicated by: ProMED-mail Rapporteur Brent Barrett

[2]

Date: Sat 21 Apr 2007

Source: Mailman School of Public Health, Columbia University, press release (edited)

<http://www.prnewswire.com/cgi-bin/stories.pl?ACCT=104&STORY=/www/stor</pre>

Scientists Discover New Virus Responsible for Deaths of 3 Transplant Recipients From Single Donor in Victoria, Australia

Knowledge of genetic sequence of virus will enable improvements in screening to enhance transplantation safety. Scientists in the Greene Infectious Disease Laboratory of Columbia University Mailman School of Public Health and colleagues in the Victoria Infectious Diseases Reference Laboratory in Melbourne, Australia and 454 Life Sciences have discovered a new virus that was responsible for the deaths of 3 transplant recipients who received organs from a single donor in Victoria, Australia.

The previously unknown virus, which is related to lymphocytic choriomeningitis virus (LCMV), was found using rapid sequencing technology established by 454 Life Sciences and bioinformatics algorithms developed in the Greene Laboratory with support from the National Institute of Allergy and Infectious Diseases. Known strains of LCMV have been implicated in a small number of cases of disease transmission by organ transplantation [see references below], however, the newly discovered virus is sufficiently different that it could not be detected using existing screening methods.

Over 30 000 organ transplants are performed in the U.S. each year. Knowledge of the genetic sequence of this virus will enable improvements in screening that will enhance the safety of transplantation.

Ian Lipkin, MD, director of the Greene Laboratory and Principal Investigator of the Northeast Biodefense Center, emphasized the importance of academic, public health, and industrial partnership in this work. "This was a team effort. Drs. Mike Catton and Julian Druce at the Victorian Infectious Disease Reference Laboratory reached out to us after a comprehensive state-of-the-art investigation failed to turn up leads," stated Dr. Lipkin. "We succeeded in identifying the virus responsible for the deaths by building on their work and utilizing new tools for pathogen surveillance and discovery developed in the Greene Laboratory and 454 Life Sciences."

(Lymphocytic choriomeningitis virtu (LCMV) is the type species of the genus _Arenavirus_ of the _Areanviridae_ family of bipartitie genome

RNA viruses. The reservoir hosts of almost all arenaviruses are rodents. LCMV is found in wild and laboratory mice, and other related "old world" arenaviruses are found in African species of rodents. Human LCMV infection may occur in rural and urban areas with high densities of rodents. Laboratory-acquired infections occur sporadically, and, previously, there have been a small number of cases of LCMV transmission by organ transplantation as mentioned by Professor Lipkin above. The virus detected by Professor Lipkin's group appears to be an LCMV-like agent but distinct from previously isolated strains of LCMV. It is unresolved, however, whether these organ-transplanted viruses are merely passengers or are responsible also for tissue-rejection illness and death. - Mod.CP]

[see also: 2005

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LCMV, transplant recipients, fatal - USA (02) 20050526.1459 LCMV, transplant recipients, fatal - USA 20050524.1426

LCMV & birth defects - USA 19951119.1095]mpp/cp/msp/lm

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医薬品 研究報告 調査報告書

識別番号·報告回数 一般的名称 販売名(企業名)		人赤血球濃厚液		報告日	第一報入手日 2007. 4. 19	新医薬品 該当	等の区分 。 なし	機構処理欄	
					AbuBakar S, Sam IC,	S, Roslan N.			
				研究報告の公表状況	MatRahim N, Hooi PS Emerg Infect Dis. 200 Jan;13(1):147-9.				
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告の概要								血液を介するウイルス、 細菌、原虫等の感染 vCJD等の伝播のリスク	
	<u>, </u>	最告企業の意見			今後の対応				
感染/	が再興し、ウイルス	されていなかったチ のゲノム配列は他の いたとの報告である		日本赤十字社では、輸加有無を確認し、帰国後4 ニヤ感染が確認されただ チクングニヤ熱の既往歴 ている。今後も引き続き、 報の収集に努める。	週間は献血不適とし 上め、渡航歴確認の復 がある場合、治癒後	ている。国内 故底を図って 6ヵ月間は献	でチクング いる。また、 血不適とし	-5.	

