

RESEARCH

8. Gajdusek DC, Gibbs CJ Jr, Alpers M. Transmission and passage of experimental "kuru" to chimpanzees. *Science*. 1967;155:212-4.
9. Shaked GM, Shaked Y, Kariv-Inbal Z, Halimi M, Avraham I, Gabizon R. A protease-resistant prion protein isoform is present in urine of animals and humans affected with prion diseases. *J Biol Chem*. 2001;276:31479-82. DOI: 10.1074/jbc.C100278200
10. Kariv-Inbal Z, Ben-Hur T, Grigoriadis NC, Engelstein R, Gabizon R. Urine from scrapie-infected hamsters comprises low levels of prion infectivity. *Neurodegener Dis*. 2006;3:123-8. DOI: 10.1159/000094770
11. Seeger H, Heikenwalder M, Zeller N, Kranich J, Schwarz P, Gaspert A, et al. Coincident scrapie infection and nephritis lead to urinary prion excretion. *Science*. 2005;310:324-6. DOI: 10.1126/science.1118829
12. Heikenwalder M, Zeller N, Seeger H, Prinz M, Klöhn PC, Schwarz P, et al. Chronic lymphocytic inflammation specifies the organ tropism of prions. *Science*. 2005;307:1107-10. DOI: 10.1126/science.1106460
13. Reed LJ, Muench H. A simple method of estimating fifty per cent endpoints. *American Journal of Hygiene*. 1938;27:493-7.
14. Pizzi M. Sampling variation of the fifty percent end-point, determined by the Reed-Muench (Behrens) method. *Hum Biol*. 1950;22:151-90.
15. Spearman C. The method of "right and wrong cases" ("constant stimuli") without Gauss's formulae. *British Journal of Psychology*. 1908;2:227-42.
16. Elliott EJ, MacAuley C, Robins D, Rohwer RG. Working safely with transmissible spongiform encephalopathies. In: Richmond JY, Editor. *Anthology of biosafety*, vol. 7. Mundelein (IL): American Biological Safety Association; 2005.
17. Mathiason CK, Powers JG, Dahmes SJ, Osborn DA, Miller KV, Warren RJ, et al. Infectious prions in the saliva and blood of deer with chronic wasting disease. *Science*. 2006;314:133-6. DOI: 10.1126/science.1132661
18. Deleault NR, Harris BT, Rees JR, Supattapone S. Formation of native prions from minimal components in vitro. *Proc Natl Acad Sci U S A*. 2007;104:9741-6. DOI: 10.1073/pnas.0702662104
19. Hadlow WJ, Race RE, Kennedy RC. Temporal distribution of transmissible mink encephalopathy virus in mink inoculated subcutaneously. *J Virol*. 1987;61:3235-40.
20. Marsh RF, Burger D, Hanson RP. Transmissible mink encephalopathy: behaviour of the agent in mink. *J Infect Dis*. 1969;120:713-9.
21. Brown P, Gibbs CJ Jr, Rodgers-Johnson P, Asher DM, Sulima MP, Bacote A, et al. Human spongiform encephalopathy: the National Institutes of Health series of 300 cases of experimentally transmitted disease. *Ann Neurol*. 1994;35:513-29. DOI: 10.1002/ana.410350504
22. Sisó S, González L, Jeffrey M, Martin S, Chianini F, Steele P. Prion protein in kidneys of scrapie-infected sheep. *Vet Rec*. 2006;159:327-8.
23. Ligios C, Cancedda GM, Margalith I, Santucci C, Madau L, Macstrale C, et al. Intracutaneous and interstitial deposition of pathological prion protein in kidneys of scrapie-affected sheep. *PLoS One*. 2007;2:e859. DOI: 10.1371/journal.pone.0000859
24. Haimir AN, Kunkle RA, Miller J. M, Hall SM. Abnormal prion protein in ectopic lymphoid tissue in a kidney of an asymptomatic white-tailed deer experimentally inoculated with the agent of chronic wasting disease. *Vet Pathol*. 2006;43:367-9. DOI: 10.1354/vp.43-3-367
25. Brown P, Rohwer RG, Dunstan BC, MacAuley C, Gajdusek DC, Drohan WN. The distribution of infectivity in blood components and plasma derivatives in experimental models of transmissible spongiform encephalopathy. *Transfusion*. 1998;38:810-6. DOI: 10.1046/j.1537-2995.1998.38998408999.x
26. Head MW, Koverianou E, Taylor L, Green A, Knight R. Evaluation of urinary PrP^{Sc} as a diagnostic test for sporadic, variant, and familial CJD. *Neurology*. 2005;64:1794-6. DOI: 10.1212/01.WNL.0000161842.68793.8A
27. Narang HK, Dagdanova A, Xie Z, Yang Q, Chen SG. Sensitive detection of prion protein in human urine. *Exp Biol Med (Maywood)*. 2005;230:343-9.
28. Krause CH, Eastick K, Ogilvie MM. Real-time PCR for mumps diagnosis on clinical specimens—comparison with results of conventional methods of virus detection and nested PCR. *J Clin Virol*. 2006;37:184-9. DOI: 10.1016/j.jcv.2006.07.009
29. Rota PA, Khan AS, Durigon E, Yuran T, Villamarzo YS, Bellini WJ. Detection of measles virus RNA in urine specimens from vaccine recipients. *J Clin Microbiol*. 1995;33:2485-8.
30. Tonry JH, Brown CB, Cropp CB, Co JK, Bennett SN, Nerurkar VR, et al. West Nile virus detection in urine. *Emerg Infect Dis*. 2005;11:1294-6.
31. Georgsson G, Sigurdsson S, Brown P. Infectious agent of sheep scrapie may persist in the environment for at least 16 years. *J Gen Virol*. 2006;87:3737-40. DOI: 10.1099/vir.0.82011-0
32. Johnson CJ, Phillips KE, Schramm PT, McKenzie D, Aiken JM, Pedersen JA. Prions adhere to soil minerals and remain infectious. *PLoS Pathog*. 2006;2:e32. DOI: 10.1371/journal.ppat.0020032
33. Seidel B, Thomzig A, Buschmann A, Groschup MH, Peters R, Beckes M, et al. Scrapie agent (strain 263K) can transmit disease via the oral route after persistence in soil over years. *PLoS One*. 2007;2:e435. DOI: 10.1371/journal.pone.0000435

Address for correspondence: Robert G. Rohwer, Veterans Affairs Medical Center, Research Service, 10 North Greene St, Mailstop 151, Baltimore, MD 21201, USA; email: rohwer@umaryland.edu

EMERGING INFECTIOUS DISEASES *online*

www.cdc.gov/eid

To receive tables of contents of new issues send an email to listserv@cdc.gov with subscribe eid-toc in the body of your message.

医薬品 研究報告 調査報告書

識別番号・報告回数		報告日	第一報入手日 2008. 8. 18	新医薬品等の区分 該当なし	機構処理欄
一般的名称	新鮮凍結人血漿	研究報告の公表状況	Bishop MT, Ritchie DL, Will RG, Ironside JW, Head MW, Thomson V, Bruce M, Manson JC. PLoS ONE. 2008 Aug 6;3(8):e2878.	公表国 英国	
販売名(企業名)	新鮮凍結血漿「日赤」(日本赤十字社) 新鮮凍結血漿-LR「日赤」(日本赤十字社)				
研究報告の概要	<p>○輸血による二次感染後のvCJD病原体に重大な変化はない</p> <p>背景:輸血による変異型クロイツフェルトヤコブ病(vCJD)の伝播が同定されたことから、輸血経路による二次伝播後のvCJD病原体に変化があるかを調べることにした。病原性や宿主適応性が増加すれば、vCJDの重大な二次アウトブレイク発生の可能性に関するリスク分析の再評価が必要である。一般集団にvCJDキャリアがいる可能性が高いため、輸血や汚染された外科用器具などの経路から更に感染する可能性がある。</p> <p>方法:我々は、野生マウスおよびトランスジェニックマウスに、輸血関連vCJD感染第1号症例由来材料を接種した。</p> <p>主な知見:輸血関連vCJD感染株の伝播の特性は、ウシ海綿状脳症(BSE)からの伝播に関連したvCJD株と著しい類似を見せている。</p> <p>結論:ヒトにおいて第2の感染経路を通してBSE病原体が適応することにより、ヒトに対する病原性の毒性が増加し、その後の伝播リスクがより大きくなるという仮説が立てられたが、本稿に示した2匹のマウスモデルのデータからは、vCJDのヒト-ヒト伝播後の病原体の伝播効率に重大な変化はないことが示される。</p>				<p>使用上の注意記載状況・ その他参考事項等</p> <p>新鮮凍結血漿「日赤」 新鮮凍結血漿-LR「日赤」</p> <p>血液を介するウイルス、 細菌、原虫等の感染 vCJD等の伝播のリスク</p>
	報告企業の意見	<p>野生マウスおよびトランスジェニックマウスに、輸血関連vCJD感染第1号症例由来材料を接種したところ、輸血による二次感染後のvCJD病原体に重大な変化はないことが明らかになったとの報告である。</p>			

No Major Change in vCJD Agent Strain after Secondary Transmission via Blood Transfusion

Matthew T. Bishop¹, Diane L. Ritchie¹, Robert G. Will¹, James W. Ironside¹, Mark W. Head¹, Val Thomson², Moira Bruce², Jean C. Manson^{2*}

¹ National CJD Surveillance Unit, University of Edinburgh, Western General Hospital, Edinburgh, United Kingdom, ² Roslin Institute, Neuropathogenesis Division, University of Edinburgh, Edinburgh, United Kingdom

Abstract

Background: The identification of transmission of variant Creutzfeldt-Jakob disease (vCJD) by blood transfusion has prompted investigation to establish whether there has been any alteration in the vCJD agent following this route of secondary transmission. Any increase in virulence or host adaptation would require a reassessment of the risk analyses relating to the possibility of a significant secondary outbreak of vCJD. Since there are likely to be carriers of the vCJD agent in the general population, there is a potential for further infection by routes such as blood transfusion or contaminated surgical instruments.

Methodology: We inoculated both wild-type and transgenic mice with material from the first case of transfusion associated vCJD infection.

Principal Findings: The strain transmission properties of blood transfusion associated vCJD infection show remarkable similarities to the strain of vCJD associated with transmission from bovine spongiform encephalopathy (BSE).

Conclusions: Although it has been hypothesized that adaptation of the BSE agent through secondary passage in humans may result in a greater risk of onward transmission due to an increased virulence of the agent for humans, our data presented here in two murine models suggest no significant alterations to transmission efficiency of the agent following human-to-human transmission of vCJD.

Citation: Bishop MT, Ritchie DL, Will RG, Ironside JW, Head MW, et al. (2008) No Major Change in vCJD Agent Strain after Secondary Transmission via Blood Transfusion. PLoS ONE 3(8): e2878. doi:10.1371/journal.pone.0002878

Editor: Corinne Ida Lasmezas, The Scripps Research Institute, United States of America

Received: May 8, 2008; **Accepted:** July 9, 2008; **Published:** August 6, 2008

Copyright: © 2008 Bishop et al. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Funding: This study was funded by the UK Department of Health and the Medical Research Council. The sponsors had no role in study design, data analysis, or the decision to submit for publication.

Competing Interests: The authors have declared that no competing interests exist.

* E-mail: Jean.Manson@roslin.ed.ac.uk

Introduction

Variant Creutzfeldt-Jakob disease (vCJD) is an acquired form of human transmissible spongiform encephalopathy (TSE) caused by infection by the bovine spongiform encephalopathy (BSE) agent that entered the human food chain in the United Kingdom during the 1980s and early 1990s. [1,2] 164 cases of vCJD have been identified in the United Kingdom and a further 41 cases in other countries worldwide. Annual mortality rates indicate that the vCJD outbreak is now in decline in the UK following a peak in 1999/2000. [3] In 2003 the first case of human-to-human secondary transmission of vCJD via blood transfusion was identified through a collaborative study between the UK National Blood Services, the National CJD Surveillance Unit, and the Office of National Statistics (Transfusion Medicine Epidemiology Review, TMER). [4,5] Statistical analysis showed that the possibility of this case being due to BSE infection was in the order of 1:15,000 to 1:30,000. [4] This patient had received a transfusion of non-leucodepleted red cells that had originated from a donor who 3 years 4 months later developed clinical vCJD. The blood recipient was methionine homozygous at codon 129 of the

prion protein (PrP) gene (*PRNP*), the same genotype as all tested vCJD cases. [6]

Two further cases of vCJD linked to blood transfusion, in MM genotype individuals, have subsequently been identified through the TMER study. [7,8] Following the discovery of these cases policy changes were made in relation to blood donation in the UK and elsewhere. In 2004 the UK Blood Service deferred transfusion recipients from acting as blood donors.

A fourth case, of asymptomatic infection following blood transfusion, was described in 2004 and this individual was heterozygous (MV) at codon 129. [9] This case was the first indication that individuals with *PRNP* genotypes other than MM could be infected by the vCJD agent. All three codon 129 genotypes are now thought to be susceptible to vCJD infection following the identification of two VV genotype appendix tissues positive for vCJD associated PrP (PrP^{Sc}) in an anonymous screening study, and the successful transmission of vCJD to 'humanised' transgenic mice of each genotype. [10–12]

The implications of these findings are that a significant number of the UK population may be carriers of vCJD infectivity, that some of the individuals may be donating blood, and that not only

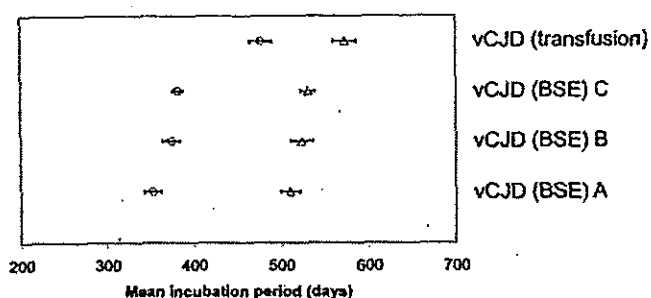


Figure 1. Comparison of incubation periods in wild-type mice. Incubation period plot comparison of vCJD (transfusion) case versus transmissions in wild-type mice of vCJD (BSE) from three sources. (Data shows mean incubation period \pm standard error of the mean. Open circles RIII line and open triangles VM line.)
doi:10.1371/journal.pone.0002878.g001

those with an MM genotype may be susceptible to infection from this source. Our research in transgenic models indicates that MV and VV individuals are likely to remain in an infectious preclinical state for a significant period of time with incubation periods potentially longer than average lifespan. [12] The identification of four instances of secondary transmission of vCJD infection from a group of 66 individuals known to have received blood products from vCJD donors, including only 28 who survived at least five years post transfusion indicates that blood transfusion is a significant risk factor for vCJD. This is likely to be due to either the route of transmission being more efficient or the agent being more infectious on human-to-human transmission or a combination of both.

TSE transmission by the blood transfusion route has been investigated in a sheep model. [13,14] These studies used intravenous (i.v.) transfusion of whole blood and blood fractions from clinical and preclinical sheep infected with BSE or scrapie. Preliminary data showed that the i.v. route gave relatively short and consistent incubation periods suggesting an efficient transmission route, with success rates of 60% for sheep infected with BSE and 40–45% for natural scrapie. [14,15]

Strain characterisation using a standard panel of inbred lines of wild-type mice originally demonstrated that BSE and vCJD agents had similar biological properties following transmission. [2,16] Similar work in other murine models has also been undertaken to study other human TSEs (genetic and iatrogenic CJD [17], and

sporadic CJD [2]), and has been used to examine emerging TSEs (atypical BSE [18] and chronic wasting disease in deer and elk [19]). [20] The development of transgenic mice expressing human PrP has led to further dissection of the nature of human TSE strains, including transmission of vCJD to gene targeted human transgenic mice. [12,17,21,22] Extensive data from studies in both wild-type and transgenic models at the Neuropathogenesis Division provide an essential background which will allow us to identify any change in the transmission characteristics of vCJD following secondary transmission. [2,12,23]

To investigate the nature of the transmissible agent following secondary transmission from human-to-human following blood transfusion we have examined the biological properties of brain material from the first case of transfusion-associated vCJD inoculated into panels of both wild-type, and transgenic mice expressing human PrP.

Results

Clinical signs of a TSE in the transgenic mice were rare and occurred after long incubation periods (IP) as found in our previous study. [12] Inoculation of the vCJD (transfusion) case produced one clinically positive HuMM mouse (at 659 days post inoculation), two positive HuMV mice (at 596 and 638 dpi) and no positive HuVV mice. Transmission of the vCJD (transfusion) case to the RIII and VM lines showed extended incubation periods compared to the three vCJD (BSE) cases. However, the hierarchy of incubation periods in the two wild-type lines was identical. (Figure 1 and Table 1) These data also show close similarities to previously published vCJD (BSE) transmission to wild-type mice despite different methodologies. These earlier studies used cerebellar material for the inoculum which was injected by simultaneous intracerebral and intraperitoneal routes. [2,23,24]

The frequency of transgenic mice positive for TSE associated vacuolation was similar between the vCJD (transfusion) case and the published vCJD (BSE) case [12], with positive results in 8/15 HuMM, 0/17 HuMV, and 0/17 HuVV mice and 6/16 HuMM, 1/15 HuMV, and 1/15 HuVV mice respectively. Regional distribution of TSE vacuolation in the brain was assessed through lesion profiling. All wild-type and the HuMM transgenic lines had sufficient positive mice to generate a profile ($n \geq 6$ mice). The overall pattern of the lesion profiles was the same in the vCJD (transfusion) and vCJD (BSE) cases for all lines of mice, however,

Table 1. Clinical and pathological assessment of wild-type mice.

Inoculum	Mouse Line	Mice Inoculated ^a	Positive for Clinical TSE Signs	Positive for TSE Vacuolation	Incubation Period (days \pm SEM)
vCJD(BSE) A	RIII	20	12	12	352.76 \pm 9.78
vCJD(BSE) B	RIII	20	18	17	374.35 \pm 9.98
vCJD(BSE) C	RIII	21	17	16	381.88 \pm 6.07
vCJD (transfusion)	RIII	23	18	18	477.33 \pm 12.68
vCJD(BSE) A	VM	22	15	22	510.20 \pm 10.97
vCJD(BSE) B	VM	22	20	21	523.75 \pm 12.57
vCJD(BSE) C	VM	21	13	18	530.69 \pm 8.16
vCJD (transfusion)	VM	22	15	18	572.90 \pm 12.96

Wild-type mouse lines RIII and VM, inoculated with vCJD(BSE) and vCJD(transfusion) were assessed clinically and pathologically for signs of TSE and mean incubation periods calculated.

^aThe group of 24 was reduced due to unavailability of some brain material for analysis.

doi:10.1371/journal.pone.0002878.t001

for the former case the VM and HuMM mice scores were lower. (Figure 2)

Immunocytochemical (ICC) detection of disease associated abnormal PrP in paraffin sections was also used as a method of assessing whether mice were transmission positive. There were 13/14 HuMM, 8/17 HuMV, 1/17 HuVV positive mice in the vCJD (transfusion) case, which was similar to the frequency of positives in the published vCJD (BSE) case: 11/15 HuMM, 11/13 HuMV, 1/15 HuVV mice. ICC data can be used to show variation in targeting of abnormal PrP deposition in the brain and variation in the nature of deposits. The ICC pattern in transgenic mice inoculated with the vCJD (transfusion) case matched that reported for vCJD (BSE) [12]. The thalamus was specifically targeted with deposition of abnormal PrP, and for the HuMM mice the hippocampus contained many intensely stained plaques including vCJD transmission associated florid plaques. ICC pattern in wild-type mice also showed similarities between the data sets with abnormal PrP deposition targeted to the thalamus and hippocampus, and large aggregates in the white matter of the corpus callosum. (Figure 3)

Biochemical analysis of disease-associated PrP by Western blot can discriminate between human cases of vCJD and sporadic CJD. [25] In the vCJD (transfusion) case the HuMM mice had a type 2B gel mobility and glycoform ratio identical to that found in vCJD (BSE) transmission to HuMM mice, and in vCJD itself. (Figure 4) Brain tissue from both vCJD (transfusion) [4] and published vCJD (BSE) [26] patients showed the type 2B pattern. The levels of PrP^{Sc} seen in the HuMV and HuVV were too low to allow typing by this standard Western blot method.

Discussion

Secondary passage of vCJD infection via blood transfusion in an MM codon 129 genotype individual results in a clinical disease phenotype and pathological characteristics that are similar to vCJD derived from BSE. [4] In this paper we confirm that the agent strain properties of primary and secondary vCJD cases are similar in transmission studies in transgenic and wild-type mice. Strain characteristics can be assessed by the frequency of clinical signs in recipient animals, the incubation period, neuropathological features, and PrP typing. All these parameters were similar in the transmission studies of primary and secondary vCJD in transgenic mice, indicating that the strain properties of the vCJD agent have not changed significantly following secondary passage in humans.

There were some differences in the results of the transmission studies which deserve further comment. The incubation period in wild-type mice was relatively extended in the vCJD(transfusion) case. However, the hierarchy of incubation periods in different inbred mouse strains was unchanged and the most plausible explanation for these findings is that, rather than implicating a change in agent characteristics, the titre of infectivity was less in the brain sample from the vCJD(transfusion) case. The distribution and degree of vacuolation was identical in the RIII mice. (Figure 2) While the distribution was identical in the VM and HuMM mice the degree of vacuolation intensity was lower for the vCJD(transfusion) case. This variability could be due to the much longer incubation times observed in these lines of mice or due to minor changes of the strain properties.

Preliminary investigation of the individuals diagnosed with vCJD following blood transfusion does not indicate a change in the neuropathological characteristics of vCJD following secondary transmission, although further studies are required to confirm this observation.

The level of infectivity in peripheral tissues in secondary cases of vCJD is unknown, although spleen and a lymph node were PrP positive in the sub-clinical case linked to blood transfusion. Evidence from BSE inoculation of primates indicates similar peripheral distribution of disease associated PrP following either oral or intravenous infection. [27] Further studies are required to assess the anatomical distribution, strain properties and level of infectivity in peripheral tissues in secondary vCJD infection. This may be important for accurate assessment of the public health risks associated with the potential for iatrogenic transmission of vCJD, which are not solely defined by the agent characteristics in brain.

Blood transfusion appears to be a relatively efficient means of secondary transmission of vCJD. To date, there have been four such transmissions in a cohort of 28 individuals who survived at least five years following transfusion of blood derived from individuals incubating vCJD. Despite extensive exposure of the UK population to the BSE agent in the food chain, there have been a relatively limited number of primary cases of vCJD (164 in the UK) and the outbreak has been in decline since 1999/2000. An important question is why there should be a disparity in the apparent efficiency of infection between primary and secondary vCJD. Transmission is generally more efficient within species than between species which may explain this observation. [28,29] Inoculation of wild-type mice with material from primary and secondary BSE passage in macaques showed that the BSE agent retained a characteristic lesion profile even though the second passage incubation period in the macaques was reduced by 50%. [30] This suggests that efficiency of transmission may increase without obvious changes to the agent strain.

Another factor is that the intravenous route of infection is very much more efficient than the oral route, as shown in experimental models. [27,31,32] Results from this study suggest the major factor here is likely to be the route of infection rather than any changes in the strain of agent. Future studies, including those using experimental oral exposure to infectivity in transgenic mice, will further address this issue.

All the primary and secondary clinical cases of vCJD have occurred in individuals with a MM genotype. The sub- or pre-clinical transfusion related infection was in a codon 129 heterozygote and genotyping of positive appendix samples identified in a screening study confirmed valine homozygosity in 2 of 3 samples tested. [10] This indicates that individuals with all codon 129 genotypes may be infected with the vCJD agent and the effect of the MV or VV background on the characteristics of the vCJD agent have not been addressed by the data in this paper.

In conclusion, transmission studies indicate that the strain characteristics of vCJD have not been significantly altered by secondary transmission through blood transfusion. This suggests that the risk of onward transmission of vCJD through other routes, for example contaminated surgical instruments, have not been increased by adaptation of the infectious agent to humans following secondary passage. However the characteristics of the infectious agent in different genetic backgrounds has not yet been defined and the prevalence of vCJD infection in the general population remains uncertain. There is need to continue to implement appropriate policies to protect against the risk of secondary transmission of vCJD until many of the remaining uncertainties are resolved.

Materials and Methods

The transgenic mice (HuMM, HuMV, HuVV) used in these experiments have been described previously. [12] These mice express human PrP under the regulation of the murine promoter